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Wheat Improvement

Food Security in a Changing Climate



CGIAR



International Maize and Wheat Improvement Center

OPEN ACCESS



Springer

Chapter 3

Defining Target Wheat Breeding Environments



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Abstract The main objective of a plant breeding program is to deliver superior germplasm for farmers in a defined set of environments, or a target population of environments (TPE). Historically, CIMMYT has characterized the environments in which the developed germplasm will be grown. The main factors that determine when and where a wheat variety can be grown are flowering time, water availability and the incidence of pests and diseases. A TPE consists of many (population) environments and future years or seasons, that share common variation in the farmers' fields, it can also be seen as a variable group of future production environments. TPEs can be characterized by climatic, soil and hydrological features, as well as socioeconomic aspects. Whereas the selection environments (SE) are the environments where the breeder does the selection of the lines. The SE are identified for predicting the performance in the TPE, but the SE may not belong to the TPE. The utilization of advanced statistical methods allows the identification of GEI to obtain higher precision when estimating the genetic effects. Multi-environmental testing (MET) is a fundamental strategy for CIMMYT to develop stable high grain yielding germplasm in countries with developing economies. An adequate MET strategy allows the evaluation of germplasm in stress hotspots and the identification of representative and correlated sites; thus, breeders can make better and targeted decisions in terms of crossing, selection and logistic operations.

Keywords Mega-environments (ME) · Target population of environments (TPEs) · Selection environments (SEs) · Genotype-by-environment interaction (GEI)

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3.1 Learning Objectives

- Identify the factors that drive wheat adaptation for the classification of target environments.
- Identify the statistical methods that can be used for defining TPEs.
- Identify the importance of a multi-environmental testing strategy for wheat breeding with a global scope.

3.2 Introduction: Wheat Mega-environments in History and the Context of Global Wheat Breeding

The success of a breeding program and, particularly, of a program with international dimensions such as CIMMYT's wheat breeding program, depends heavily on the characterization of the environments where the germplasm will be grown.

Historically, since its earliest efforts to breed wheat in the 1940s, CIMMYT has characterized the environments in which the developed germplasm will be grown. At that time, this characterization was restricted to the geographical areas in Mexico. However, as soon as CIMMYT's mandate became global in the 1970s, 15 agroecological zones were defined, for instance, the region that encompasses India, Pakistan, Bangladesh and Nepal (South Asia), or the Nile Valley zone (Egypt and Sudan).

Those agroecological zones were redefined in the late 1980s. As it became evident that the sole geographical description was inadequate, given the diversification of the production systems, the need for high-yielding and stable germplasm and the simple fact that for specific conditions, certain traits were needed, particularly those related to stress tolerance and quality requirements. Hence, this redefinition led to the concept of Mega-Environment (ME), described by Rajaram et al. [1] as a "broad" but not necessarily "contiguous area" with similar biotic and abiotic constraints, cropping systems, consumer preferences and production volume. By 1992, twelve ME had been conceived, six for each spring and winter growth habit. Here we present only those corresponding to spring wheat (Table 3.1).

After CIMMYT's target environment classification, and thanks to the historical data collected by the International Wheat Improvement Network (IWIN), this was followed by several reports on germplasm adaptation and performance in the context of ME (see Chap. 7). A study published by DeLacy et al. [4] demonstrated that the major discrimination factors for these ME were latitude and the presence/absence of stresses, plus the agreement between the ME-based locations and the location groups obtained from pattern analysis. Another study reported by Hodson and White [3] defined ME classification with the aid of GIS tools, and hence, a more quantitative and specific classification was proposed. In such a study, it was demonstrated that long term environmental variables, mainly temperature and precipitation in the coolest and wettest quarter of the year, were effective to separate environments based on abiotic stresses and growth habits, *i.e.*, spring vs. winter growth type.

Table 3.1 Spring bread wheat mega-environments, land area, characteristics and required traits

Mega environment	Area (ha) ^a	Climate conditions ^b	Biotic stresses	Abiotic stresses	Key agronomic traits
1	47.2	Favorable, irrigated, low rainfall. Coolest quarter mean min temp ≥ 3 °C < 11 °C	Rusts (leaf, yellow and stem rust)	Lodging	Water and nutrient use efficiency
2	5.9	a. High rainfall. Highland summer rain. Wettest quarter (three consecutive wettest months) mean min temp ≥ 3 °C < 16 °C, wettest quarter precipitation ≥ 250 mm, elevation ≥ 1400 m b. High rainfall. Lowland winter rain. Coolest quarter mean min temp ≥ 3 °C < 16 °C, coolest quarter precipitation ≥ 150 mm, elevation < 1400 m	Yellow rust, stem rust, STB & FHB	Lodging	Sprouting resistance
3	1.3	Same as ME2, topsoil pH < 5.2	Same as ME2	Acid soils	
4	13.5	a. Low rainfall. Coolest quarter mean min temp ≥ 3 °C < 11 °C, wettest quarter precipitation ≥ 100 mm < 400 mm c. Low rainfall, stored moisture. Coolest quarter mean min temp ≥ 3 °C < 16 °C, wettest quarter precipitation ≥ 100 mm < 400 mm	Rusts, STB, tan spot, root diseases	Drought	Water use efficiency
5	2.1	Coolest quarter mean min temp > 11 °C < 16 °C, High rainfall or irrigated	Rusts and spot blotch in low rainfall areas, Rusts and fusarium in high rainfall areas	High temperature	Early maturity
6	21	High latitude (> 45 °N or S). Coolest quarter mean min temp less than -13 °C, warmest quarter mean min temp ≥ 9 °C	Rusts	High temperature and drought	Photosensitivity

^aData from Lantican et al. [2]; ^bClimatic conditions according to Hodson and White [3]

Additional studies derived from the historical data provided by the IWIN have shown how CIMMYT wheat germplasm performs and adapts throughout the locations within each ME [5–7]. These studies also demonstrated that CIMMYT’s main yield testing site located in northwest Mexico correlates positively with the locations that belong to each ME for spring wheat.

3.3 Major Factors That Broadly Impact the Definition of Target Environments

3.3.1 Flowering Time: Photoperiod and Vernalization

Flowering time is a fundamental adaptive trait, as it determines where and when a variety can be grown, and, in general, largely determines the reproductive success of a plant. Flowering must occur during an optimal environmental period that permits the full development of the reproductive organs. This period should also be long enough to allow optimal grain filling.

One factor that highly determines flowering time in wheat is photoperiod. In wheat there is a series of dominant genes (*Ppd*) located on chromosomes 2A, 2B and 2D that induce an insensitive reaction to photoperiod [8–10]. Photoperiod insensitivity means that plants reach flowering even under short days, provided that any vernalization requirements have been met. One characteristic of the wheat cultivars derived from the Green Revolution is their insensitivity to photoperiod, which along with their short stature and disease resistance, significantly contributed to their adaptation to a broad range of environments.

Various studies have been conducted to determine the advantages of photoperiod-sensitive (PS) and photoperiod-insensitive germplasm (PI). For high latitude locations, evidence indicates that PS germplasm may have an advantage over PI germplasm [11, 12]. High GEI in Northern Europe [11], North America [13], and other high latitude locations in Asia [14], indicates that regional adaptation plays a major role in breeding spring wheat.

The geographical division suggested by Worland et al. [11] in Europe where PI and PS spring wheat germplasm displays better adaptation is 45–46° N. For practical purposes, wheat grown north of Paris is frequently PS, while south of that latitude the germplasm that better adapts is PI due to the summer conditions in Southern Europe [11].

Another factor that largely determines flowering time is *vernalization*. In this context, vernalization is the exposure to cold temperatures after germination to acquire or accelerate the ability to flower [15]. In northern latitudes where winters are cold, vernalization sensitivity is required, as it delays floral initiation which consequently protects ear development when low temperatures can severely damage it [16], hence conferring adaptability to northern latitudes.

The distinction between spring and winter growth habits is determined by a series of genes that can express both sensitivity and insensitivity to vernalization. Of these series of genes—*Vrn1*, *Vrn2* and *Vrn3*—*Vrn1* and *Vrn3* on the homoeologous groups 5 and 7, respectively, are dominant for spring growth habit, while *Vrn2* on chromosome 5A is dominant for winter growth habit [17]. In winter wheat sown and germinated in the autumn, *Vrn2* suppresses *Vrn3*, which in turn impedes the expression of *Vrn1*; then, as winter approaches, lower temperatures downregulate *Vrn2*, facilitating the upregulation of *Vrn3*, which in turn promotes *Vrn1* transcription for the induction of flowering [18].

In geographical regions where wheat is grown during the winter and harvested late in the spring, the presence of *Vrn1* dominant genes confers adaptability to those lower latitude regions. The *Vrn-A1* and *Vrn-D1* genes of the *Vrn1* series are the most common, although all three (*Vrn-A1*, *Vrn-B1* and *Vrn-D1*) are present in CIMMYT's germplasm either alone or in combination [19].

The two previously mentioned factors—photoperiod and vernalization—alone, broadly determine the target breeding environments (high and low latitude regions) and, consequently, the type of germplasm that is required for each environment/s, since they guideline the planting and harvesting times.

The paradigm until the early years (1940s) of wheat breeding in Mexico dictated that breeding must be conducted in the environment where the future varieties will be cultivated [20]. However, given the need to accelerate the development of high-yielding and stem rust resistant germplasm, two generations per year started to be grown—using shuttle breeding—with the sole objective of speeding up the breeding process (Chap. 30 describes new technologies to speed up breeding). This paradigm shift took place years before any deep knowledge on the photoperiod in/sensitivity in wheat was available [21]. As germplasm exchange happened through the assembling of the first international yield trials during 1960s, the daylength effect on the materials became evident, since those shuttled-bred wheats developed in Mexico would adapt in most places in latitudes lower than 45° N [22].

3.3.2 *Water Availability and Temperature*

Water availability for the wheat crop is paramount to determine key traits in breeding. Water availability can favor optimal growing conditions, in the absence of high temperatures. However, drought stress is a constraint for wheat production in locations where water access is limited, either because of the lack of irrigation equipment or because the climate is dry (low rainfall).

Drought is one of the most severe factors that reduce wheat productivity (see Chap. 23 for details). In meteorological terms, it is defined as the absence of rain for a certain period, during which plants suffer from the lack of water in the soil. Yield losses of 20% can occur if plants are grown with 40% less water than required to avoid the stress [23]. This loss varies depending on the phenological stage at which

the stress occurs, for instance, it can be larger if water is limited at the reproductive stage than if it occurs only at the vegetative stage [23].

Plants are drought stressed when water for the roots is limited and when the transpiration rate becomes higher. Drought can affect germination and plant establishment, growth, biomass accumulation, leaf senescence and, consequently, grain yield, but at the cellular level, it affects membrane integrity, pigment content, osmotic adjustment, photosynthetic activity, gas exchange and cell elongation [24].

Regions that are typically considered prone to drought stress are North Africa, some regions in West and Central Asia, and some locations in South America. Regions that are considered optimal in terms of water availability are the Nile Valley in Africa, the Northwestern Gangetic Plains and Northwestern Mexico.

Temperature is considered a stress factor that drastically influences wheat productivity once vernalization requirements—if any—have been met (see Chap. 22). Temperatures above optimum thresholds take high relevance, particularly in the context of climate change, since it determines the traits that the plants must carry to cope with the stress, such as earliness to avoid terminal heat stress [25]. It is estimated that for every °C increase above the a base temperature (13 °C) grain yield decreases by 6% [26]. Higher temperatures modify wheat phenology by reducing the number of days to reach flowering and maturity, consequently reducing the number of days in which plants can intercept light for photosynthesis, which leads to a reduction in biomass and grain yield. Larger yield reductions are expected in tropical and subtropical regions where wheat is grown, such as regions in India, which is a major wheat producer in the world [27].

3.3.3 Diseases

Following the fundamental paradigm in plant pathology (disease triangle), a disease outbreak occurs if there is (1) an adequate (susceptible) host, (2) a virulent pathogen, and if (3) favorable environmental conditions are present (see Chap. 19 for details). Hence, diseases tend to follow specific distribution patterns depending on the whether their environmental requirements are met.

While rusts, as a group of diseases, are found in all wheat growing areas, other leaf diseases occur in certain environments and crop management conditions (see Chaps. 8, 9 and 19 for details). Disease distribution and occurrence are dependent on both temporal and spatial variation, and these factors determine the resistance traits that cultivars must carry for certain environments. For instance, tan spot (caused by *Pyrenophora tritici-repentis*) incidence is linked to an expansion of zero-tillage practices (in Brazil, Argentina, Paraguay) or in places where climate does not allow fast stubble decomposition (Central Asia) and monocropping is common [28]. Septoria tritici blotch (caused by *Mycosphaerella graminicola*) is most common in temperate (15–20 °C) and humid wheat growing regions. Powdery mildew (caused by *Blumeria graminis* f. sp. *tritici*) is common in highly productive

areas with maritime or semi-continental climate, particularly in China and South America [28].

3.4 Target Population of Environments

The main objective of a plant breeding program is to deliver superior germplasm for farmers in a defined set of environments, or a target population of environments (TPE). A TPE consists of many (population) environments and future years or seasons. A TPE is also a variable group of future production environments. Climatic (seasonal) variation in the same farmer's field might change drastically year after year causing the exacerbation of GEI. GEI can have two components: (1) static and predictable (repeated) variability due to the location (site) where the trial has been established, and (2) dynamic and unpredictable variability due to the year effect.

Target environments should be characterized by climatic, soil and hydrological characteristics as well as by socioeconomic characteristics. There are different ways to group trials and environments into TPE. One is to group together sites where line means are highly correlated. A standard methodology is to use *stratified hierarchical cluster analyses* of the sites based on climatic variables and production traits [29].

The selection environments (SE) are the environments where the breeder does the selection of the lines. The SE are identified for predicting the performance in the TPE, but the SE may not belong to the TPE. If the lines in the SE predict those in the TPE, then (1) it is important to compute the genetic correlations between the lines in the SE versus the same (or related lines) in the TPE and show some relatively high correlations between lines in SE and in TPE; (2) for screening lines in the SE, the repeatability (broad-sense heritability) in the SE should be high; (3) SE should allow a large number of lines to be screened at a low cost, so SE should allow high selection intensity (i).

3.5 Multi-environmental Testing and Genotype-by-Environment Interactions

As CIMMYT's mandate became international, the observations made between 1944 and the 1960s established the bases for the definition of target environments on a global scale. Along with this, the implementation of a breeding strategy based on ME targeted breeding, a diverse gene pool for crossing, shuttle breeding, selection under optimal conditions and multilocation testing have led to the enhancement of the adaptability and stability that characterize CIMMYT spring wheat germplasm to date.

Multi-environmental testing (MET) is a paramount strategy for CIMMYT to develop stable high grain yielding germplasm in countries with developing

economies (see Chap. 7). An adequate MET strategy allows the evaluation of germplasm in stress hotspots and the identification of representative and correlated sites; thus, breeders can make better and targeted decisions in terms of crossing, selection and logistic operations. Another highly important aspect for CIMMYT's MET strategy is that collaborators can directly evaluate CIMMYT's elite germplasm, and hence they can make line selections for further evaluation and variety release, as well as utilize the germplasm as parental lines in their breeding programs to improve local adaptation.

Every year CIMMYT undertakes significant efforts to distribute international nurseries that comply with global and local seed health regulations, to collaborators within the IWIN, with the only request of returning the data to CIMMYT, for breeders to analyze them in a global context and support breeding decisions. The nurseries are of three different types: yield trials, observation nurseries (prior to yield trials), and trait specific nurseries (Table 3.2), Chap. 7 describes in detail international yield trials for bread wheat. Between 2013 and 2017, CIMMYT's wheat germplasm was distributed to 350 collaborators in 80 countries per year.

Despite the large variability between MEs, it is possible to simulate the most significant ones at CIMMYT's main testing site in northwest Mexico, Ciudad Obregon, a semi-arid location with suitable infrastructure for irrigation and available machinery for establishing the planting systems common around the globe. At this location in the Yaqui Valley, it is possible to mimic optimal, drought and heat stressed environments by applying the water management system corresponding to each ME, in combination with different sowing dates. This MET at one single location that is highly correlated with representative international locations [6, 7, 30] allows the breeders to select the elite germplasm that will most likely have an outstanding performance in international yield trials, and will consequently provide National Agricultural Research Centers a selection of CIMMYT's best materials every year.

Analysis of these international trials requires the utilization of advanced statistical methods that are able to parsimoniously model the GEI, obtain higher precision when estimating the genetic effects, and allow the identification of GEI patterns, for instance, the Factor Analytic (FA) and Sites Regression (SREG) models [31–33]. The FA model utilizes the leading principal components of the GEI covariance matrix in a mixed model framework, and accounts for the maximum amount of variation with a reduced number of parameters [32]. In the SREG model, the genotype and the GEI are estimated together, which is useful for evaluating METs, as its first and second principal components account for the non-crossover and the crossover interaction, respectively [34]. This property allows a visual examination to discriminate genotypes and sites with and without crossover interactions [34]. The FA and SREG models have been used to identify the trend of genetic gains and site correlations in CIMMYT's international nurseries [30, 35].

Table 3.2 International nurseries annually distributed by CIMMYT within the International Wheat Improvement Network

Nursery type	Trial/nursery	Abbreviation	Target environment	Grain Color	BW/DW ^a
Yield trials	Elite Spring Wheat Yield Trial	ESWYT	ME1	White	BW
	Harvest Plus Yield Trial	HPYT	ME1	White	BW
	Heat Tolerant Wheat Yield Trial	HTWYT	ME5	White	BW
	High Rainfall Wheat Yield Trial	HRWYT	ME2	Red	BW
	High Rainfall Wheat Screening Nursery	HRWSN	ME2	Red	BW
	Int. Durum Yield Nursery	IDYN	ME1, ME4, ME5		DW
	South Asia Bread Wheat Genomic Prediction Yield Trial	SABWGPYT	ME1, ME4, ME5	White	BW
Observation	Int. Bread Wheat Screening Nursery	IBWSN	ME1, ME4, ME5	White	BW
	Int. Durum Screening Nursery	IDSN	ME1, ME4, ME5		DW
	Semi Arid Wheat Screening Nursery	SAWSN	ME4	White	BW
	Semi Arid Wheat Yield Trial	SAWYT	ME4	White	BW
	Wheat Yield Consortium Yield Trial	WYCYT			BW
		Fusarium Head Blight Screening Nursery	FHBSN		
Trait specific	Harvest Plus South Asia Screening Nursery	HPAN	ME1, ME4, ME5	White	BW
	Heat Tolerance Screening Nursery	HTSN	ME5	White/Red	BW
	Helmithosporium Leaf Blight Screening Nursery	HLBSN			
	Int. Septoria Observation Nursery	ISEPTON			BW
	Karnal Bunt Screening Nursery	KBSN		White	BW
	Stem Rust Resistance Screening Nursery	SRRSN		White/Red	BW
	Stress Adaptive Trait Yield nursery	SATYN			BW

^aBW Bread Wheat, DW Durum wheat

3.6 Example of TPE Definition

We applied the mentioned methodology to a set of locations in India with data from the Elite Spring Wheat Yield Trials that are distributed internationally by CIMMYT, upon request. Daily meteorological data for these locations in India were obtained

from the NASA Langley Research Center POWER Project funded through the NASA Earth Science/Applied Science Program. Then we implemented a principal component analysis to infer the number of groups (TPEs) that would explain most of the variation and then perform hierarchical clustering with the Euclidean distance matrix of data. From our analysis we obtained three main TPEs for India, in agreement with the wheat producing zones determined by the Indian government (Table 3.3 and Fig. 3.1), and that together account for more than 97% of India's wheat producing area. Finally, we obtained the correlated response to selection,

Table 3.3 Agroecological zones for wheat production in India and CIMMYT's breeding target population of environments (TPE)

Zone	Area covered	Estimated area (m ha)	Estimated productivity (kg/ha)	Estimated production (mt)	TPE
Northern Hills Zone (NHZ)	Western Himalayan regions of Jammu & Kashmir (except Jammu & Kathua dist.); Himachal Pradesh (except Una & Paonta Valley); Uttarakhand (except Tarai area); Sikkim & hills of West Bengal & North Eastern States	0.82	2203	1.81	
North Western Plains Zone (NWPZ)	Punjab, Haryana, Delhi, Rajasthan (except Kota & Udaipur divisions), western Uttar Pradesh (except Jhansi division), parts of Jammu & Kashmir (Jammu & Kathua District), parts of Himachal Pradesh (Una district & Paona valley) and Uttarakhand (Tarai region)	12.33	4527	55.82	TPE1
North Eastern Plains Zone (NEPZ)	Eastern Uttar Pradesh, Bihar, Jharkhand, Odisha, West Bengal, Assam and plains of North Eastern States	8.85	2509	22.20	TPE2
Central Zone (CZ)	Madhya Pradesh, Chhattisgarh, Gujrat, Rajasthan (Kota & Udaipur divisions), Uttar Pradesh (Jhansi division)	6.84	2978	20.37	TPE3
Peninsular Zone (PZ) & Southern Hills Zone (SHZ)	PZ: Maharashtra, Karnatka, Andhra Pradesh, Telengana, Goa, plains of Tamil Nadu. SHZ: Hilly areas of Tamil Nadu & Kerla comprising the Nilgiri & Palni hills of southern plateau	0.71	1404	1.00	TPE3
All Zones		29.55	3424	101.20	

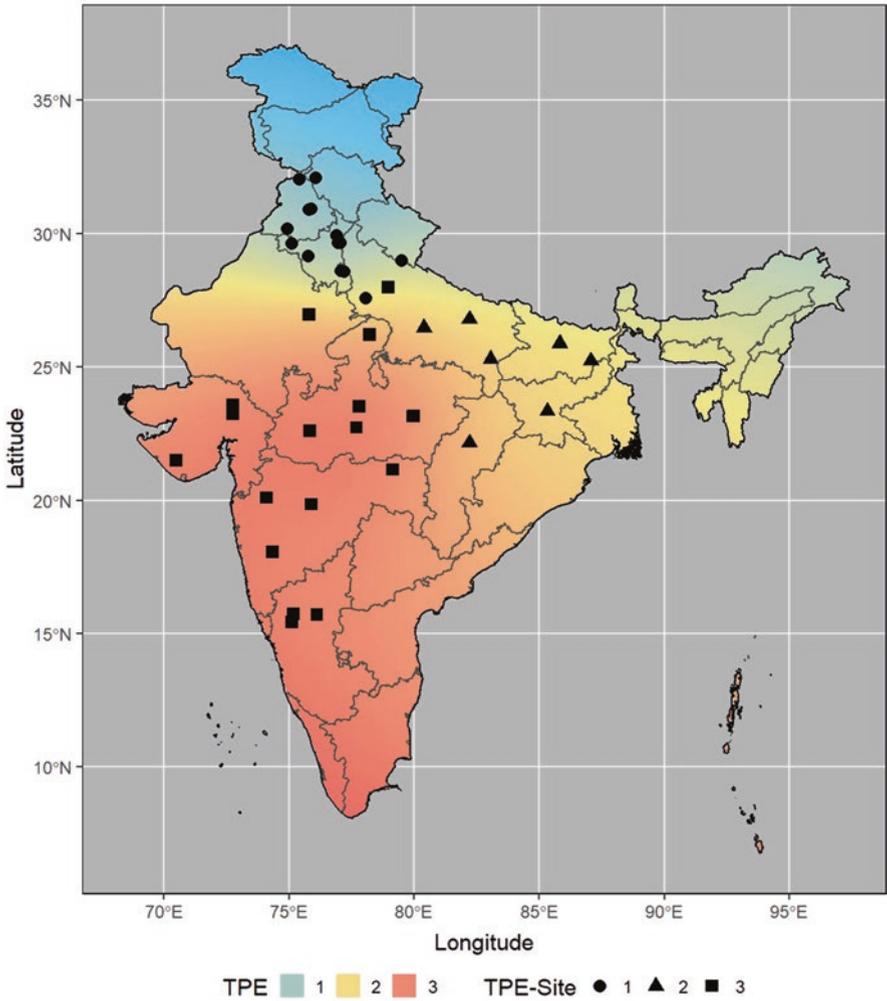


Fig. 3.1 TPE classification in India, obtained from environmental data

between each TPE and SE in Mexico, by first obtaining the genetic correlations with Eq. 3.1.

$$r_A = \frac{p_{i,j}}{\sqrt{h_i^2 * h_j^2}} \tag{3.1}$$

where r_A is the genetic correlation, $p_{1,2}$ represents the phenotypic correlation between site i and j , and h_i^2 and h_j^2 are the heritability of sites i and j , respectively.

Then, assuming that the same selection intensity applied in the SE is applied in the TPE (Eq. 3.2):

$$CR = \bar{r}_A * \sqrt{\frac{h_{SE}^2}{h_{TPE}^2}} \quad (3.2)$$

where CR is the correlated response to selection, \bar{r}_A is the genetic correlation averaged over years of testing, h_{SE}^2 is the heritability of the SE and h_{TPE}^2 is the heritability of the TPE.

Our results indicate that the centralized breeding efforts in combination with the MET can give a selection efficiency (CR) as high as in the TPE, assuming the same selection intensity (Table 3.4). However, several factors are in place to obtain this result: CIMMYT's main yield testing site allows the simulation of various environments, the high heritability usually observed in the testing phase, a relatively stable (semi-arid) climate with favorable temperatures, water availability, irrigation infrastructure and mechanized operations. Furthermore, this result does not consider the fact that the selection intensity can be several times higher in the SE than in the TPE, given that several thousands of lines (~9000) are tested annually under optimal conditions (Stage 1 testing), from which ~1000 are selected to be tested in the SE (Stage 2), and ~300 are evaluated in the SE in the Stage 3 of testing, to finally distribute 46–48 new elite lines in each international yield trial nursery (Table 3.2).

Table 3.4 Average heritability (H^2), genetic correlations and correlated selection response between SE and TPE in India

		Average H^2	Genetic correlation			Correlated selection response		
			1	2	3	1	2	3
SE^a	B2IR	0.62	0.40	0.62	0.45	0.46	1.05	0.63
	B5IR	0.61	0.63	0.55	0.31	0.73	0.93	0.43
	BLHT	0.85	0.32	0.54	0.25	0.43	1.08	0.42
	F5IR	0.65	0.51	0.50	0.40	0.61	0.87	0.56
	FDRP	0.59	0.23	0.10	0.34	0.26	0.16	0.47
TPE	1	0.46						
	2	0.21						
	3	0.32						

^aSelection environments (SE) are: Beds 5 irrigations (B5IR): Trials conducted on raised beds with full irrigation management (optimal), 500 mm of available water. Flat 5 irrigations (F5IR): Trials planted on flat land with full irrigation (optimal), 500 mm of available water. Beds 2 irrigations (B2IR). Trials conducted on raised beds with partial irrigation, 260 mm of available water. Flat Drip (FDRIP): Trials planted on flat land with severe drought, 180 mm of available water. Beds late heat (BLHT): Trials planted in February, subject to heat stress and fully irrigated, 500 mm of available water

3.7 Key Concepts and Conclusions

Characterizing TPEs is critical for any plant breeding endeavor to succeed. Determining the main factors that may limit wheat productivity in a determined set environments (TPEs) is fundamental to incorporate key traits in breeding. Such limitations include, but are not limited to: water availability, temperature and incidence of pests and diseases. Additionally, for a breeding program to succeed, it is important that the SE display relatively high correlations with the TPEs, allow a higher selection intensity and accuracy of selection (higher repeatability). At CIMMYT's main testing location in northwest Mexico, it is possible to mimic optimal, drought and heat stressed environments to artificially create SE that, at one single location, are highly correlated with representative international locations to allow breeders the selection of elite germplasm with potential outstanding performance in international yield trials, and in so doing, to provide National Agricultural Research Centers a selection of CIMMYT's best materials every year.

For a refined definition of TPEs, statistical methods such as the FA model and SREG coupled with the climatic, soil, hydrological and socioeconomic characteristics of the environments can be applied to allow the identification of GEI patterns. These models (FA and SREG) have the advantage of being parsimonious and allow to measure the extend of non-crossover and crossover GEI.

Multi-environmental testing is paramount to identify high yielding and climate reliance germplasm, as well as to determine the GEI patterns that conform potential TPEs. The CIMMYT MET strategy has the benefit of evaluating the germplasm in stress hotspots, identification of representative and correlated sites, rapid response to new constraints (see Chap. 9 for examples) and direct access to germplasm for CIMMYT collaborators, so the materials can be used as parents or directly released as varieties.

References

1. Rajaram S, Van Ginkel M, Fischer RA (1994) CIMMYT's wheat breeding mega-environments (ME). In: Li ZS, Xin ZY (eds) Proceedings of the 8th International wheat genetic symposium. Beijing, pp 1101–1106
2. Lantican MA, Braun H-J, Payne TS, Singh RP, Sonder K, Michael B, van Ginkel M, Erenstein O (2016) Impacts of international wheat improvement research, pp 1994–2014
3. Hodson DP, White JW (2007) Use of spatial analyses for global characterization of wheat-based production systems. *J Agric Sci* 145:115
4. DeLacy IH, Fox PN, Corbett JD, Crossa J, Rajaram S, Fischer RA, van Ginkel M (1993) Long-term association of locations for testing spring bread wheat. *Euphytica* 72:95–106. <https://doi.org/10.1007/BF00023777>
5. Braun H-J, Pfeiffer WH, Pollmer WG (1992) Environments for selecting widely adapted spring wheat. *Crop Sci* 32:1420–1427. <https://doi.org/10.2135/cropsci1992.0011183X003200060022x>

6. Lillemo M, Van Ginkel M, Trethowan RM, Hernandez E, Rajaram S (2004) Associations among international CIMMYT bread wheat yield testing locations in high rainfall areas and their implications for wheat breeding. *Crop Sci* 44:1163–1169
7. Lillemo M, Van Ginkel M, Trethowan RM, Hernandez E, Crossa J (2005) Differential adaptation of CIMMYT bread wheat to global high temperature environments. *Crop Sci* 45:2443–2453
8. Mohler V, Lukman R, Ortiz-Islas S, William M, Worland AJ, Van Beem J, Wenzel G (2004) Genetic and physical mapping of photoperiod insensitive gene Ppd-B1 in common wheat. *Euphytica* 138:33–40. <https://doi.org/10.1023/B:EUPH.0000047056.58938.76>
9. Beales J, Turner A, Griffiths S, Snape JW, Laurie DA (2007) A pseudo-response regulator is misexpressed in the photoperiod insensitive Ppd-D1a mutant of wheat (*Triticum aestivum* L.). *Theor Appl Genet* 115:721–733. <https://doi.org/10.1007/s00122-007-0603-4>
10. Bentley AR, Turner AS, Gosman N, Leigh FJ, Maccaferri M, Dreisigacker S, Greenland A, Laurie DA (2011) Frequency of photoperiod-insensitive Ppd-A1a alleles in tetraploid, hexaploid and synthetic hexaploid wheat germplasm. *Plant Breed* 130:10–15. <https://doi.org/10.1111/j.1439-0523.2010.01802.x>
11. Worland AJ, Borner A, Korzun V, Li WM, Petrovic S, Sayers EJ (1998) The influence of photoperiod genes on the adaptability of European winter wheats. *Euphytica* 100:385–394. <https://doi.org/10.1023/a:1018327700985>
12. Dyck JA, Matus-Cádiz MA, Hucl P, Talbert L, Hunt T, Dubuc JP, Nass H, Clayton G, Dobb J, Quick J (2004) Agronomic performance of hard red spring wheat isolines sensitive and insensitive to photoperiod. *Crop Sci* 44:1976–1981. <https://doi.org/10.2135/cropsci2004.1976>
13. Lanning SP, Hucl P, Pumphrey M, Carter AH, Lamb PF, Carlson GR, Wichman DM, Kephart KD, Spaner D, Martin JM, Talbert LE (2012) Agronomic performance of spring wheat as related to planting date and photoperiod response. *Crop Sci* 52:1633–1639. <https://doi.org/10.2135/cropsci2012.01.0052>
14. Trethowan RM, Morgunov A, He Z, De Pauw R, Crossa J, Warburton M, Baytasov A, Zhang C, Mergoum M, Alvarado G (2006) The global adaptation of bread wheat at high latitudes. *Euphytica* 152:303–316. <https://doi.org/10.1007/s10681-006-9217-1>
15. Chouard P (1960) Vernalization and its relations to dormancy. *Annu Rev Plant Physiol*. <https://doi.org/10.1146/annurev.pp.11.060160.001203>
16. Law CN, Worland AJ (1997) Genetic analysis of some flowering time and adaptive traits in wheat. *New Phytol* 137:19–28. <https://doi.org/10.1046/j.1469-8137.1997.00814.x>
17. Kamran A, Iqbal M, Spaner D (2014) Flowering time in wheat (*Triticum aestivum* L.): a key factor for global adaptability. *Euphytica* 197:1–26. <https://doi.org/10.1007/s10681-014-1075-7>
18. Distelfeld A, Li C, Dubcovsky J (2009) Regulation of flowering in temperate cereals. *Curr Opin Plant Biol* 12:178–184
19. Beem J, Mohler V, Lukman R, Ginkel M, William M, Crossa J, Worland AJ (2005) Analysis of genetic factors influencing the developmental rate of globally important CIMMYT wheat cultivars. *Crop Sci* 45:2113–2119. <https://doi.org/10.2135/cropsci2004.0665>
20. Borlaug NE (1983) Contributions of conventional plant breeding to food production. *Science* 219(80):689–693
21. Mckinney HH, Sando WJ (1935) Earliness of sexual reproduction in wheat as influenced by temperature and light in relation to growth phases. *J Agric Res* 5:621–541
22. Rajaram S (1994) Wheat germplasm improvement: historical perspectives, philosophy, objectives, and missions. In: Rajaram S, Hettel GP (eds) *Wheat breeding at CIMMYT: commemorating 50years of research in mexico for global wheat improvement*. CIMMYT, Texcoco, pp 1–10
23. Daryanto S, Wang L, Jacinthe P-A (2016) Global synthesis of drought effects on maize and wheat production. *PLoS One* 11:e0156362. <https://doi.org/10.1371/journal.pone.0156362>
24. Anjum SA, Xie X, Wang L, Saleem MF, Man C, Lei W (2011) Morphological, physiological and biochemical responses of plants to drought stress. *Afr J Agric Res* 6:2026–2032. <https://doi.org/10.5897/ajar10.027>

25. Mondal S, Singh RP, Crossa J, Huerta-Espino J, Sharma I, Chatrath R, Singh GP, Sohu VS, Mavi GS, Sukuru VSP, Kalappanavar IK, Mishra VK, Hussain M, Gautam NR, Uddin J, Barma NCD, Hakim A, Joshi AK (2013) Earliness in wheat: a key to adaptation under terminal and continual high temperature stress in South Asia. *Field Crop Res* 151:19–26. <https://doi.org/10.1016/j.fcr.2013.06.015>
26. Asseng S, Ewert F, Martre P, Rotter RP, Lobell DB, Cammarano D, Kimball BA, Ottman MJ, Wall GW, White JW, Reynolds MP, Alderman PD, Prasad PVV, Aggarwal PK, Anothai J, Basso B, Biernath C, Challinor AJ, De Sanctis G, Doltra J, Fereres E, Garcia-Vila M, Gayler S, Hoogenboom G, Hunt LA, Izaurrealde RC, Jabloun M, Jones CD, Kersebaum KC, Koehler A-K, Muller C, Naresh Kumar S, Nendel C, O'Leary G, Olesen JE, Palosuo T, Priesack E, Eyshir Rezaei E, Ruane AC, Semenov MA, Shcherbak I, Stockle C, Stratonovitch P, Streck T, Supit I, Tao F, Thorburn PJ, Waha K, Wang E, Wallach D, Wolf J, Zhao Z, Zhu Y (2015) Rising temperatures reduce global wheat production. *Nat Clim Chang* 5:143–147
27. Asseng S, Cammarano D, Basso B, Chung U, Alderman PD, Sonder K, Reynolds M, Lobell DB (2017) Hot spots of wheat yield decline with rising temperatures. *Glob Chang Biol* 23:2464–2472. <https://doi.org/10.1111/gcb.13530>
28. Duveiller E, Singh RP, Nicol JM (2007) The challenges of maintaining wheat productivity: pests, diseases, and potential epidemics. *Euphytica* 157:417–430. <https://doi.org/10.1007/s10681-007-9380-z>
29. Cooper M, Hammer GL (1996) Plant adaptation and crop improvement. CAB International, Wallingford
30. Crespo-Herrera LA, Crossa J, Huerta-Espino J, Autrique E, Mondal S, Velu G, Vargas M, Braun HJHJ, Singh RPRP (2017) Genetic yield gains in CIMMYT's international elite spring wheat yield trials by modeling the genotype \times environment interaction. *Crop Sci* 57:789–801. <https://doi.org/10.2135/cropsci2016.06.0553>
31. Burgueño J, Crossa J, Cornelius PL, Yang R-C (2008) Using factor analytic models for joining environments and genotypes without crossover genotype \times environment interaction. *Crop Sci* 48:1291. <https://doi.org/10.2135/cropsci2007.11.0632>
32. Meyer K (2009) Factor-analytic models for genotype \times environment type problems and structured covariance matrices. *Genet Sel Evol* 41:21. <https://doi.org/10.1186/1297-9686-41-21>
33. Crossa J, Vargas M, Joshi AK (2010) Linear, bilinear, and linear-bilinear fixed and mixed models for analyzing genotype \times environment interaction in plant breeding and agronomy. *Can J Plant Sci* 90:561–574. <https://doi.org/10.4141/CJPS10003>
34. Crossa J, Vargas M, Cossani CM, Alvarado G, Burgueño J, Mathews KL, Reynolds MP (2015) Evaluation and interpretation of interactions. *Agron J* 107:736. <https://doi.org/10.2134/agnonj2012.0491>
35. Crespo-Herrera L, Crossa J, Huerta-Espino J, Vargas M, Mondal S, Velu G, Payne TSS, Braun H, Singh RPP (2018) Genetic gains for grain yield in CIMMYT's semi-arid wheat yield trials grown in suboptimal environments. *Crop Sci* 58:1890–1898. <https://doi.org/10.2135/cropsci2018.01.0017>

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